



Review

Role of the interleukin 6 receptor family in epithelial ovarian cancer and its clinical implications



Janani Kumar, Alister C. Ward*

School of Medicine, Deakin University, Geelong, Victoria, Australia

Strategic Research Centre in Molecular and Medical Research, Deakin University, Geelong, Victoria, Australia

ARTICLE INFO

Article history:

Received 7 June 2013

Received in revised form 8 December 2013

Accepted 24 December 2013

Available online 2 January 2014

Keywords:

Epithelial

Ovarian cancer

IL-6R

Cytokine signaling

JAK–STAT pathway

ABSTRACT

Ovarian cancer is the most lethal gynecological malignancy, with few effective treatment options in most cases. Therefore, understanding the biology of ovarian cancer remains an important area of research in order to improve clinical outcomes. Cytokine receptor signaling through the Janus kinase–signal transducer and activator of transcription (JAK–STAT) pathway is an essential component of normal development and homeostasis. However, numerous studies have implicated perturbation of this pathway in a range of cancers. In particular, members of the IL-6R family acting via the downstream STAT3 transcription factor play an important role in a number of solid tumors – including ovarian cancer – by altering the expression of target genes that impact on key phenotypes. This has led to the development of specific inhibitors of this pathway which are being used in combination with standard chemotherapeutic agents. This review focuses on the role of IL-6R family members in the etiology of epithelial ovarian cancer, and the application of therapies specifically targeting IL-6R signaling in this disease setting.

© 2014 Elsevier B.V. All rights reserved.

Contents

1.	Introduction	118
2.	Biology of ovarian cancer	118
2.1.	Types of ovarian cancer	118
2.2.	Epithelial ovarian cancer progression	118
2.3.	Immune responses to epithelial ovarian cancer	118
2.4.	Cytokines in epithelial ovarian cancer	118
3.	The IL-6R family	119
3.1.	Structure and function	119
3.2.	Downstream signaling	119
4.	Role of the IL-6R family in epithelial ovarian cancer	119
4.1.	IL-6R	119
4.2.	IL-11R	120
4.3.	LIFR	120
4.4.	OSMR	120
4.5.	IL-12R	120
4.6.	IL-23R	121
4.7.	G-CSFR	121
4.8.	OBR	121
5.	Targeting IL-6R family signaling in epithelial ovarian cancer	121
5.1.	Pathway inhibitors	121
5.1.1.	IL-6 and IL-6R antibodies	121
5.1.2.	JAK1/2–STAT3 inhibitors	121

Abbreviations: ECM, extracellular matrix; EGF, epidermal growth factor; EOC, epithelial ovarian cancer; G-CSF, granulocyte colony-stimulating factor; GP130, glycoprotein 130; IL, interleukin; JAK, Janus kinase; LIF, leukemia inhibitory factor; MAPK, mitogen-activated protein kinase; OBR, obesity receptor; OSM, oncostatin M; PI 3-K, phosphatidylinositol 3-kinase; R, receptor; SOCS, suppressor of cytokine signaling; STAT, signal transducer and activator of transcription

* Corresponding author at: School of Medicine, Deakin University, Pigdons Road, Geelong, Victoria 3216, Australia. Tel.: +61 3 5227 2041; fax: +61 3 5227 2945.

E-mail address: award@deakin.edu.au (A.C. Ward).

5.2. Cytokine therapies in ovarian cancer	122
6. Conclusions	122
Acknowledgements	122
References	122

1. Introduction

Ovarian cancer is the most lethal gynecological cancer, with approximately 200,000 new cases diagnosed each year globally, and a greater than 60% mortality rate within five years [1]. This poor prognosis is partly due to the largely asymptomatic nature of the disease, such that the primary tumor has often already spread at the time of diagnosis [2]. However, there is also a limited understanding of disease etiology at the molecular level, which continues to hamper targeted therapeutic development.

2. Biology of ovarian cancer

2.1. Types of ovarian cancer

Ovarian cancer is not a single disease, but rather a spectrum of malignancies exhibiting distinct biology and histopathology. Ovarian cancers are grouped into three major categories based on the cells from which they originate [3,4]. Germ cell tumors are derived from oocytes, and represent approximately 10%–15% of all ovarian cancers [5]. Stromal (or 'Sex cord') tumors are formed from cells of the stromal mesenchyme/sex-cord within the ovary, and may contain gonad-related cells, as well as immature fibroblasts and connective tissue-forming cells [6]. These represent about 5% of all ovarian cancers with tumors arising from granulosa cells being the most prevalent type [5]. However, the most common form of this disease is epithelial ovarian cancer (EOC), representing about 80% of all cases. EOC consists of distinct histological subtypes: predominantly serous (cells resembling the internal lining of the fallopian tube), endometrioid (cells resembling endometrium), clear cell (hobnailed shaped cells with clear cytoplasm) and mucinous (cells resembling endocervical epithelium) [7–9]. Serous is the most common subtype, representing around 70% of EOC [10]. Each EOC subtype is further divided into benign, borderline and malignant, based on the level of containment, and graded from I to IV, depending on the extent of differentiation [3], with some specific genetic alterations identified [11,12]. EOC, especially the predominant serous subtype, is mainly thought of as originating from the malignant transformation of the ovarian surface epithelium (OSE) [13] (Fig. 1). However, some researchers have argued that the key site is the fallopian tube, which shares a common embryonic origin to the OSE [14,15].

2.2. Epithelial ovarian cancer progression

During the transformation process, EOC cells acquire additional properties, such as increased cell proliferation, survival and migration. There are specific changes in cell–cell and cell–extracellular matrix (ECM) interactions, with increased proteolysis and ECM degradation observed. This culminates in the shedding of tumor cells into the peritoneum, where they survive in an anchorage-independent manner as cellular aggregates or spheroids, until they attach to a suitable secondary site for further growth [16,17] (Fig. 1). There is also a growing consensus that EOC, like many other cancers, is a stem cell disease in which a small population of cells has acquired the ability to both self-renew as well as generate the cell types that are characteristic of the tumor [18].

Women with EOC typically present with advanced-stage disease, when the cancer has already spread throughout the peritoneum, and in some cases, to distant metastatic sites. With aggressive surgery

supplemented with paclitaxel and platinum-based chemotherapy, most patients initially return to a state of microscopic disease with minimal residual tumor. However, this is usually short-lived, with recurrent disease characterized by increased metastatic behavior, as well as acquisition of drug-resistance to multiple types of chemotherapy via increased activity of the ATP-binding cassette (ABC) drug transporters, enhanced DNA repair and reduced apoptosis [19]. Only ~30% of patients with recurrent disease survive for >5 years [4].

2.3. Immune responses to epithelial ovarian cancer

The clinical outcome of EOC is strongly dependent on the immune response [20], which is critically dependent on macrophages, the most prevalent immune cell within the EOC microenvironment. 'M1' macrophages elicit strong anti-tumor immunity, whereas 'M2' macrophages suppress adaptive immunity, and promote tumor survival, invasion and metastasis [21]. Tumor-associated macrophages (TAMs) are mostly identified as being of the M2 type, with their density correlating with poor prognosis in EOC [22]. With regard to other immune cells, higher numbers of infiltrating T cells are associated with increased survival [23], with a better prognosis with a Th1 response compared to a Th2 response [24]. Increased levels of Th17 cells have also been observed in ovarian cancer [25], but these may in fact enhance tumorigenesis [26]. Tolerogenic dendritic cells are able to suppress T cell effector functions to promote cancer development [27,28].

2.4. Cytokines in epithelial ovarian cancer

Cytokines are small polypeptides released from cells to regulate the activities of other cells via interactions with specific cytokine receptors expressed on their surface [29]. They are able to stimulate proliferation, differentiation, survival and activation [7]. At least 16 different cytokines are expressed in normal ovaries, along with many of the corresponding

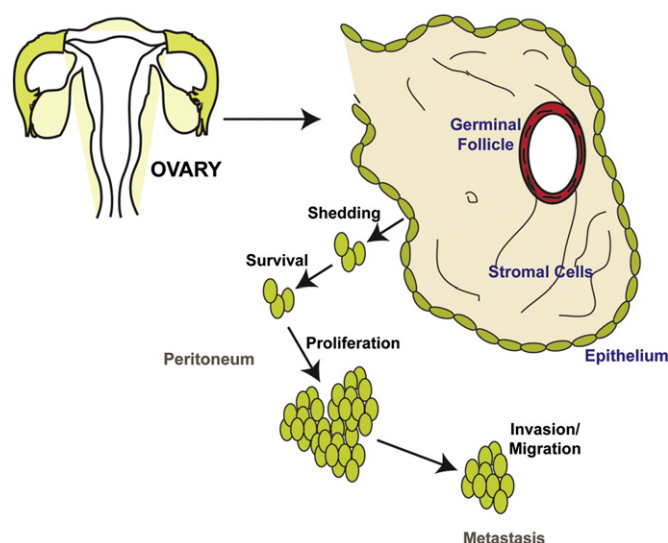


Fig. 1. Biology of ovarian cancer and its progression. Diagrammatic representation of the cell origins of ovarian cancers: germinal, stromal and epithelial. The progression of epithelial ovarian cancer is demonstrated in more detail, including the role of some key cancer phenotypes, which include proliferation, survival, invasion/migration, angiogenesis and chemoresistance.

receptors [30]. In ovarian cancer there is increased expression of particular cytokines and/or receptors [31]. These act – along with growth factors such as EGF – to enhance key phenotypes associated with tumor progression, such as proliferation, migration and survival, including chemoresistance [17]. This can be due to direct effects on the cancer cells themselves, or via indirect effects on components of the immune system.

3. The IL-6R family

3.1. Structure and function

Cytokines that act via members of the so-called interleukin-6 receptor or “IL-6R” family of cytokine receptors (Fig. 2) are of particular relevance to EOC. The core members of this family employ a ligand-specific receptor α chain in combination with a shared receptor chain, glycoprotein 130 (GP-130), and sometimes a third receptor chain. However, also in this family are receptors that utilize GP130-related receptor chains, including granulocyte colony-stimulating factor receptor (G-CSFR) and obesity gene receptor (OBR), which form homomeric complexes [32]. Soluble forms of several receptors exist, including of the IL-6R (sIL-6R), which can bind ligand extracellularly before associating with signaling chains on target cells [33].

Individual IL-6R family members have distinct functions: for example, IL-6R mediates immune, hematopoietic and hepatic responses, and contributes to follicle development in normal ovaries [34,35], leukemia inhibitory factor receptor (LIFR) functions in stem cell maintenance, neural and hematopoietic development [36], oncostatin M receptor (OSMR) has a role in the differentiation and survival of neural and hematopoietic cells [37], G-CSFR promotes granulocytic development, hematopoietic stem cell mobilization and myeloid cell migration [38], while OBR, the receptor for leptin, functions in the regulation of energy intake and energy homeostasis [39]. The expression of these receptors and their cytokine ligands is normally tightly controlled. Overexpression of both IL-6 and its receptors (IL-6R and sIL-6R) in the serum of cancer patients has typically been associated with poor clinical outcomes [40].

3.2. Downstream signaling

Within the IL-6R family, the common utilization of GP130 or related receptor chains to transduce intracellular signals means that similar downstream pathways are activated [32]. These include the mitogen-activated protein kinase (MAPK), phosphatidylinositol 3-kinase (PI3K) and, probably most importantly, the Janus kinase–signal transducer and activator of transcription (JAK–STAT) pathways [33,41]. JAK tyrosine kinases are constitutively associated with IL-6R family members and are activated following ligand binding [33]. These phosphorylate receptor tyrosines, forming docking sites for signaling molecules including STATs, which in turn also become activated. While other JAKs and STATs may also be involved, virtually all members of the IL-6R family utilize the JAK1/2–STAT3 signaling pathway as a core mechanism for mediating their downstream effects [42]. Phosphorylation of STAT3 causes it to

dimerize and translocate from the cytoplasm to the nucleus where it initiates changes in the transcription of a number of genes, including those affecting growth, differentiation, and survival [43,44]. Although the full spectrum of STAT3 target genes is yet to be defined, several have been identified that contribute to specific phenotypes (Fig. 3) [45]. These include the pro-proliferative genes Cyclin D1 and c-MYC [46], the anti-apoptotic genes BCL-2, Survivin and MCL-1 [46], genes involved in migration/invasion, such as MMP-9, CXCR4, CXCL12 and Integrins [47,48], the pro-angiogenic genes VEGF and Notch3 [46], as well as genes mediating chemoresistance, including MDR1 and GSTpi [49]. In addition, genes encoding feedback regulators, such as suppressor of cytokine signaling (SOCS) 3 [50] are also induced, acting in combination with tyrosine phosphatases and other regulatory proteins to extinguish signaling from IL-6R family members [51].

4. Role of the IL-6R family in epithelial ovarian cancer

There is increasing evidence that signaling via specific IL-6R family members may play a significant role in the progression of various cancers, including EOC.

4.1. IL-6R

IL-6 is a potent, pleiotropic cytokine that acts via a receptor complex consisting of the ligand-binding IL-6R α chain and the signal-transducing GP130 chain, both of which are widely expressed [33].

IL-6 has been implicated in a wide range of cancers, such as multiple myeloma [52], endometrial cancer [53], lung cancer [54], colorectal cancer [55], renal cell carcinoma [56], cervical cancer [57] and breast cancer [58]. IL-6 is known to directly promote tumor cell attachment, proliferation and migration through the activation of several downstream signaling pathways, including both the JAK–STAT and MAPK cascades [40]. Other studies have shown that IL-6 can also enhance survival [58] and angiogenesis [59]. Unchecked production of IL-6 can also lead to chronic inflammation, which is implicated in many types of cancer [60].

IL-6/IL-6R signaling is important in EOC. IL-6 is constitutively secreted from the ovarian cancer microenvironment, either directly from ovarian cancer cells or through secondary inflammation [61,62]. IL-6 production is increased by EGF stimulation in advanced-stage epithelial ovarian cancer [63] and relevant cell lines [64], indicating cross-talk between growth factors and cytokine signaling. High plasma levels of IL-6 correlate with poor prognosis [65–68], and are associated with ovarian cancer subtypes with poorer outcomes [69]. Increased expression of the IL-6R has also been observed in EOC [48,64,70]. This includes soluble forms of the receptor, generated either via alternate splicing or increased expression of sheddases that cleave the full-length receptor, which can mediate effects in tumor cells as well as adjacent stromal cells or peritoneal cells [70].

IL-6/IL-6R signaling can act in a number of ways to augment ovarian cancer progression. IL-6 can act directly on ovarian cancer cells to enhance migration, attachment and invasion [48,64,71], and facilitate

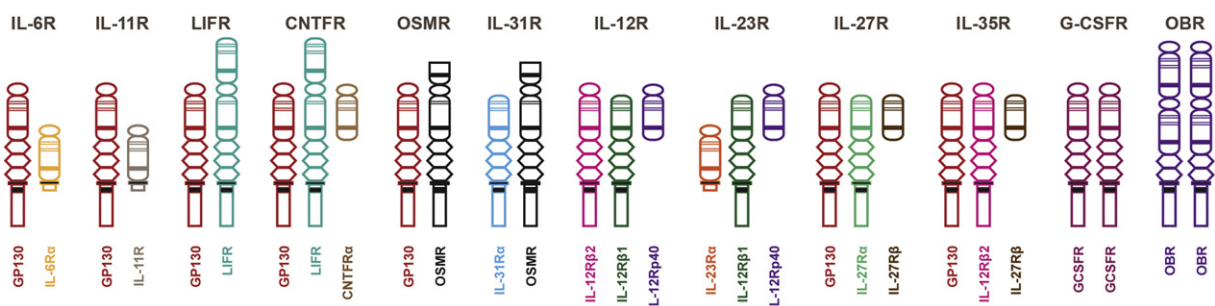


Fig. 2. The interleukin-6 receptor family. Schematic representation of the IL-6R family members, showing their constituent receptor chains, which are indicated below.

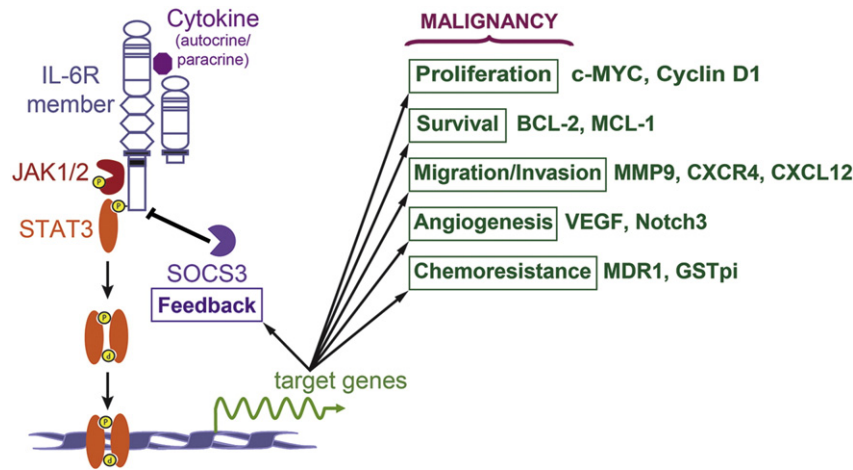


Fig. 3. IL-6R family signaling in ovarian cancer. Signaling from the IL-6R family of receptors (autocrine or paracrine) via JAK1/2–STAT3 and its impact on expression of target genes involved in the key malignant phenotypes of proliferation, survival, migration/invasion, angiogenesis and chemoresistance, as well as negative feedback.

anchorage-independent growth [48]. For example, an IL-6R-positive EOC cell line exhibited ~50% increased tumor formation in mice compared with cells lacking IL-6R expression [70]. Finally, IL-6 can enhance chemoresistance [49,72], through increased expression of multi drug resistance pumps and anti-apoptotic genes in ovarian cancer cells [49]. Consistent with this, IL-6 is observed in the serum and ascites of patients harboring resistant cancers [66,67,73,74].

IL-6 can also act indirectly, via a number of mechanisms [59,75]. Firstly, it can polarize macrophages toward the M2 phenotype [76–78]. These TAMs are able to suppress immune responses as well as stimulate angiogenesis through VEGF and IL-8 production, and promote metastasis via MMP-2 and MMP-9 [48,79]. Secondly, IL-6 can preferentially differentiate CD4⁺ T cells toward Th17 subsets [80], which can be further expanded by this cytokine within the ovarian tumor microenvironment [25]. These Th17 cells can promote tumor growth by increasing the density of the microvasculature in EOC [81], as well as inducing further IL-6 production [82]. Thirdly, IL-6 can mediate the induction of tolerogenic dendritic cells [83]. Fourthly, IL-6 can also stimulate expression of the inhibitory co-stimulatory molecule B7-H4 on both tumor and immune cells, further suppressing immunity [84]. Finally, IL-6 has been shown to blunt the potency of chemotherapeutic agents through its actions on endothelial cells [85].

4.2. IL-11R

The receptor complex for IL-11 consists of IL-11R α and GP130 [86]. IL-11R signaling supports the growth of multiple hematopoietic progenitors as well as exerting metabolic and cytoprotective effects on cells outside the hematopoietic system [87].

IL-11 has been shown to regulate adhesion and migration of chondrosarcoma [88] and endometrial cancer [89], and was recently demonstrated to be the dominant cytokine in the etiology of gastrointestinal cancer [90]. IL-11R is expressed on >90% of EOC, with expression of its ligand much less common [91]. However, the function of IL-11/IL-11R in EOC remains uncertain. This is of particular clinical relevance, since IL-11 is being developed as a treatment for chemotherapy-induced thrombocytopenia [92].

4.3. LIFR

The LIFR consists of a heterodimer of the LIFR α chain and GP130, which acts as a receptor for leukemia inhibitory factor (LIF), cardiotrophin-1 (CTF1), as well as oncostatin M (OSM) [36]. LIFR is expressed on a wide range of hematopoietic cells and cell lines

originating from bone marrow, thymus, spleen, liver, placental tissue, and peritoneum [93], with LIFR signaling shown to be important for the maintenance of embryonic stem cells, embryo implantation, neuronal survival and hematopoiesis via LIF [36], and in cardio protection via CTF-1 [94].

The functional significance of LIFR in human solid tumor cell lines is not fully understood, but LIF and LIFR has been found to be expressed in breast, kidney and prostate cancer cell lines, and can induce tumor growth in these cells in either an autocrine or paracrine manner [95,96]. Ovarian carcinomas consistently express LIFR [97], with altered expression of LIFR linked to chemoresistance [98]. Elevated LIF has also been associated with EOC [27,99], where it appears to correlate with the clinical characteristics of the tumor [99]. LIF may also act indirectly by contributing to the development of immune deficiency within the peritoneal cavity [27], as well as augmenting to the development of TAM-like cells [100].

4.4. OSMR

The mammalian OSMR complex comprises OSMR α and GP130 [101], and is expressed on many cell types, including endothelial cells, hepatic cells, lung cells and bone marrow cells [102]. OSMR signaling is able to influence the development of blood cells, liver, and specific neuronal populations [37].

OSMR has been shown to be frequently expressed in tumors, where it can activate downstream signaling pathways upon ligand stimulation [97]. OSM has been demonstrated to suppress the growth of a subset of breast and lung carcinoma cell lines [103,104], but to increase proliferation of prostate carcinoma cell lines [105]. Ovarian carcinomas consistently express OSMR and OSM [97,106], although their role in this setting is also controversial. One group has suggested that OSM can strongly stimulate proliferation [106], while another has suggested that OSM exerts a small effect on survival, but not growth [97].

4.5. IL-12R

The IL-12R consists of a heterodimer of IL-12R β 1 and IL-12R β 2, which is activated by a heterodimer of the cytokine IL-12p35 and the soluble cytokine receptor IL-12p40 [107]. IL-12 is a pro-inflammatory cytokine, with IL-12R signaling playing a central role in promoting tumor immune-surveillance, which has seen it used in various modalities to activate the immune system in cancer [108].

IL-12 has been shown to be overexpressed in EOC, with expression significantly associated with both increased overall survival [109] and

reduced disease progression [110]. This has led to IL-12 being used in the context of EOC, with a plasmid-based IL-12 therapy is in clinical trials for platinum-sensitive ovarian cancer [111]. IL-12 transduced MSCs have also been shown to inhibit ovarian carcinoma cells both in vitro and in vivo [112].

4.6. IL-23R

The IL-23R consists of the ligand-specific IL-23R α and the shared IL-12R β 1, which is activated by a heterodimer of the cytokine IL-23p19 and the soluble cytokine receptor IL-12p40 [113]. IL-23 has been shown to be pro-inflammatory, but its role in cancer is less clear [113,114]. Like IL-12, IL-23 was overexpressed in EOC, with a positive correlation for overall survival [109]. However, TNFR-mediated enhancement of EOC growth was also shown to positively correlate with levels of IL-23 and IL-23R [115]. Moreover, polymorphisms in the IL-23R gene have been associated with late stage EOC in a Chinese population, suggesting a role in disease susceptibility [116]. One mechanism of action by which IL-23R signaling might mediate these effects is through expansion of Th17 cells.

4.7. G-CSFR

G-CSF is a secreted 25 kDa glycoprotein that binds to a homomeric single-chain receptor, G-CSFR, which is closely related to GP130 [38]. G-CSFR is expressed on a range of hematopoietic cells, such as mature neutrophilic granulocytes, myeloid progenitors, hematopoietic stem cells, monocytes and lymphocytes, as well non-hematopoietic tissues, including cardiomyocytes, neuronal precursors, endothelial cells and placental tissue [117]. Its principal role is in the G-CSF-induced mobilization of hematopoietic stem cells and production of neutrophils during “emergency hematopoiesis” [118]. These properties have seen G-CSF commonly used in the clinic to increase the production of neutrophils in patients with chemotherapy-induced neutropenia [119].

The G-CSFR has previously been implicated in a diverse range of malignancies. This includes hematological cancers, particularly acute myeloid leukemia [117], but also solid tumors. For example, a significant proportion of invasive bladder carcinomas have been shown to express both G-CSF and G-CSFR, with subsequent autocrine signaling contributing to their proliferation and survival in vitro (possibly via STAT3), as well as the size of their induced tumors in vivo [120]. G-CSF has also been demonstrated to stimulate the proliferation and migration of tumor cells derived from patients with head and neck squamous cell carcinoma, which could be blocked with neutralizing antibodies. Interestingly, while G-CSFR-positive tumors grew at a slower rate, they showed increased invasion, angiogenesis and survival [121]. Functional G-CSFR complexes have also been identified on a variety of other solid tumors [122].

The role of G-CSF/G-CSFR signaling in ovarian cancer has remained controversial. Broad expression of the G-CSFR has been reported in ovarian cancer cells, with co-expression of G-CSF often observed in these cells, or in neighboring stromal tissue, meaning both autocrine and paracrine activation is possible [123–126]. However, the clinical significance of G-CSF expression remains unclear, with one study showing it was an adverse prognostic factor if part of a paracrine loop [125], but another suggesting it did not correlate with prognosis [127]. G-CSF has been shown to support proliferation in some primary ovarian cancer cells and cell lines [123,128], but in others it inhibited proliferation [128] or had no effect [125,126,129]. G-CSF was found to enhance EGF-mediated mitogenesis in the G-CSFR-positive cell line OVCAR-3 [125], suggesting possible cross-talk with growth factor receptor signaling. We have recently demonstrated a role for G-CSF/G-CSFR signaling in EOC cell migration and survival, including in response to chemotherapeutic agents [129], suggesting it may contribute to ovarian cancer progression via several mechanisms.

4.8. OBR

Leptin, a 16 kDa protein, signals via OBR [130]. Like G-CSFR, OBR signals directly as a homodimer [131], but initiates similar signaling pathways to other IL-6R family receptors, namely the JAK–STAT, MAPK and PI3K pathways [132]. However, there exist multiple forms of the OBR with different signaling properties [133]. OBR functions in the regulation of energy metabolism, appetite, bone formation and angiogenesis [39]. However, it also has important roles in cancer cell proliferation, invasion and metastasis, including of breast, prostate, hepatic and endometrial cancers [134–137]. In the case of breast cancer, there have been extensive studies on leptin-induced cell proliferation [138,139] and epithelial-to-mesenchymal transition (EMT) [140]. The reported pro-proliferative and anti-apoptotic effects of leptin in prostate and breast cancers are mediated via STAT3 activation [141,142].

The role of leptin/OBR signaling in ovarian cancer remains less clear. Both leptin and its receptor are expressed in normal ovaries [143]. Overexpression of OBR represents a poor prognostic factor for epithelial ovarian cancer [144], but the mechanistic details are limited. For example, OBR was found to be expressed in three ovarian cancer cell lines, SKOV-3, BG-1 and OVCAR-3, but only the BG-1 cell line showed an increased rate of proliferation in response to leptin [145]. However, another study showed leptin could stimulate cell proliferation and inhibit apoptosis in the OVCAR-3 cell line [146]. Recent studies have confirmed that leptin can stimulate cell growth and inhibit apoptosis in OVCAR-3 cells through increased Cyclin D1 and Mcl-1 expression, in this case via the activation of the MAPK and PI3K signaling pathways [147]. Leptin has also been shown to contribute to the activation of the estrogen receptor pathway in ovarian cancer cells, thereby indirectly promoting tumor cell growth [148]. Interestingly, while the short forms of OBR were shown to be expressed in all EOCs, expression of the signaling-competent long receptor form was more variable [145], which may explain some of the observed variation in leptin responses.

5. Targeting IL-6R family signaling in epithelial ovarian cancer

5.1. Pathway inhibitors

Aberrant activation of IL-6R family members has been reported to play a major role in tumorigenesis [40]. In addition, JAK1/2 and STAT3 have been separately implicated in cancers and other proliferative disorders [43]. As a result, a number of specific therapeutics targeting the IL-6R–JAK1/2–STAT3 pathway are in clinical trials, including for EOC (Fig. 4).

5.1.1. IL-6 and IL-6R antibodies

Antibodies have been developed that specifically target either IL-6 or the IL-6R to block their activity, and shown to inhibit tumor growth either alone or in combination with cytotoxic chemotherapies in a range of carcinomas. For example, an anti-IL-6 antibody (CNTO-328, Siltuximab) has been used in a number of phase I/II clinical trials, including myeloma, renal, breast and prostate cancer [58,149–153]. Siltuximab inhibited IL-6-induced STAT3 induction in paclitaxel-resistant ovarian cancer cells [154], and decreased IL-6 signaling, tumor growth, angiogenesis and TAM generation in a xenotransplantation model, with evidence of efficacy in ovarian cancer patients [75]. An anti-IL-6R antibody (Tocilizumab), which inhibits ligand binding, has been demonstrated to be efficacious in rheumatoid arthritis [155], and represents an additional therapeutic option for ovarian cancer therapy. Finally, a soluble GP130 fusion (sgp130Fc) that blocks IL-6 trans-signaling reduced ascites formation and enhanced taxane sensitivity of ovarian cancer xenografts [85].

5.1.2. JAK1/2–STAT3 inhibitors

JAK1/2 and STAT3 inhibitors have been developed and used in clinical trials particularly for myeloproliferative disorders and cancers

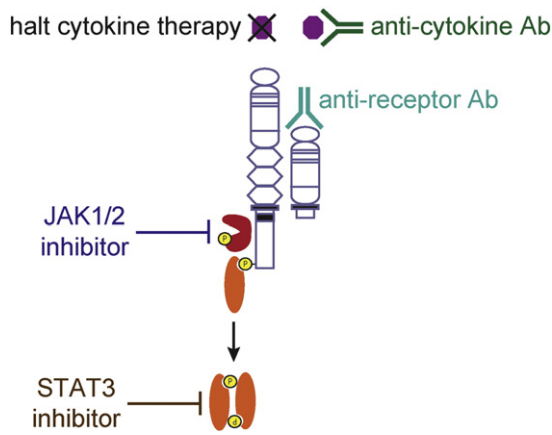


Fig. 4. Strategies for targeting IL-6R family signaling in ovarian cancer. A variety of therapeutic approaches are available to block the potential pathologic action of IL-6R family members in ovarian cancer, including cessation of cytokine therapy, antibodies to ligands/receptors, and inhibitors of the JAK1/2–STAT3 pathway.

[156–160]. Indeed, these have already shown success in ovarian tumors. Inhibition of JAK2 with diindolylmethane induced apoptosis in ovarian cancer cells [161], while the JAK inhibitor AZD1480 was able to increase the chemosensitivity of ovarian cancer cell lines by inhibiting STAT3 activation by IL-6 and oncostatin M [162]. The STAT3 inhibitor CDDO-Met was able to block IL-6 secretion and STAT3 activation in EOC cell lines, and down-regulate the anti-apoptotic proteins BCL-2 and BCL-XL in paclitaxel-resistant lines [163]. Similarly, curcumin was able to suppress both STAT3 activation and growth of EOC cells through up-regulation of the STAT3 inhibitor PIAS-3 [164]. Inhibition of STAT3 with siRNA was able to reduce growth and induce apoptosis of EOC cells [165], and reversed paclitaxel resistance in drug resistant cancer cell lines [166]. This collectively supports the notion that targeting the JAK/STAT pathway will also be efficacious in chemoresistant ovarian cancer. Recent studies have revealed the involvement of the JAK–STAT pathway in ‘cancer stem-like cells’ [167], making the use of such inhibitors even more enticing, as it may eliminate the cell population that facilitates the development of recurrent tumors. Finally, STAT3 inhibitors can indirectly reduce tumorigenicity by acting on non-cancer cells, such as decreasing angiogenesis through its effects on endothelial cell migration [43,168], or by abrogating tumor cell-mediated inhibition of DC maturation [169]. Thus, JAK1/2–STAT3 pathway inhibitors hold promise as a future therapeutic target in ovarian cancer.

5.2. Cytokine therapies in ovarian cancer

The important roles played by the IL-6R family have seen their respective cytokines become widely-used clinically. However, of particular relevance are their roles in cancer therapy. Thus, LIF has been administered to manage chemotherapy-induced peripheral neuropathy [170], IL-6 used as part of photodynamic therapy [171], while G-CSF has been extensively employed to aid hematopoietic recovery following chemotherapy for a range of cancers, including those of an ovarian nature [172]. However, if the cancer expresses the relevant receptor, these treatments may contribute to tumor development by switching on various signaling pathways that could lead to altered phenotypes, including increased proliferation, migration, survival and chemosensitivity [126]. Of particular relevance, G-CSF is commonly used in the context of ovarian cancer to restore neutrophils ablated by standard chemotherapeutic drugs used to treat patients [173]. Therefore, pre-screening patients for the presence of G-CSFR expression prior to administration of G-CSF should be considered, potentially indicating alternative approaches for these patients. Of relevance, a phase II trial showed that the co-administration of carboplatin, with granulocyte-macrophage colony stimulating factor and recombinant interferon gamma 1b in

women with recurrent ovarian cancer elicited a response rate comparable with standard approaches [174], which might be useful alternative in the case of G-CSFR positive ovarian cancer.

6. Conclusions

IL-6R family members play important roles in the etiology of epithelial ovarian cancer. Several (e.g. IL-6R, LIFR, G-CSFR) can function directly in ovarian cancer cells, enhancing proliferation, migration/invasion and/or survival, including chemoresistance. Some can act to modify immune cell responses against EOC, either in a positive (e.g. IL-12, IL-23) or negative (e.g. IL-6) manner, or can influence endothelial cells to promote angiogenesis or alter their permeability (e.g. IL-6). Downstream signaling via STAT3 plays a central role in mediating these effects. This suggests some caution should be given to the use of some potentially deleterious cytokine-based immune therapies, such as G-CSF, with screening for the presence of the cognate receptor a useful potential step forward in the cases. However, the further development of other cytokine-based approaches, such as IL-12 and IL-23, should be encouraged. Finally, the clear elucidation of common pro-tumorigenic roles provides significant opportunities to develop strategies targeting IL-6R family signaling components, including inhibitors of STAT3 activation/function, in this disease setting.

Acknowledgements

The authors recognize the support of a Deakin University International Postgraduate Scheme Award (to JK) and funding from both the Geelong Community Foundation and Atkins Charitable Trust (to ACW).

References

- [1] NBOCC, National Breast and Ovarian Cancer Centre, Global Cancer Facts and Figures, 2011.
- [2] E. Lengyel, Ovarian cancer development and metastasis, *Am. J. Pathol.* 177 (2010) 1053–1064.
- [3] V.W. Chen, B. Ruiz, J.L. Killeen, T.R. Cote, X.C. Wu, C.N. Correa, Pathology and classification of ovarian tumors, *Cancer* 97 (2003) 2631–2642.
- [4] R.A. Soslow, Histologic subtypes of ovarian carcinoma: an overview, *Int. J. Gynecol. Pathol.* 27 (2008) 161–174.
- [5] N. Auersperg, A.S. Wong, K.C. Choi, S.K. Kang, P.C. Leung, Ovarian surface epithelium: biology, endocrinology, and pathology, *Endocr. Rev.* 22 (2001) 255–288.
- [6] L.M. Roth, Recent advances in the pathology and classification of ovarian sex cord-stromal tumors, *Int. J. Gynecol. Pathol.* 25 (2006) 199–215.
- [7] A.W. Kurian, R.R. Balise, V. McGuire, A.S. Whittemore, Histologic types of epithelial ovarian cancer: have they different risk factors? *Gynecol. Oncol.* 96 (2005) 520–530.
- [8] R.T. Marquez, K.A. Baggerly, A.P. Patterson, J. Liu, R. Broaddus, M. Frumovitz, E.N. Atkinson, D.I. Smith, L. Hartmann, D. Fishman, A. Berchuck, R. Whitaker, D.M. Gershenson, G.B. Mills, R.C. Bast Jr., K.H. Lu, Patterns of gene expression in different histotypes of epithelial ovarian cancer correlate with those in normal fallopian tube, endometrium, and colon, *Clin. Cancer Res.* 11 (2005) 6116–6126.
- [9] D.A. Bell, Origins and molecular pathology of ovarian cancer, *Mod. Pathol.* 18 (Suppl. 2) (2005) S19–S32.
- [10] S.A. Cannistra, Cancer of the ovary, *N. Engl. J. Med.* 351 (2004) 2519–2529.
- [11] G. Breedlove, C. Busenhardt, Screening and detection of ovarian cancer, *J. Midwifery Womens Health* 50 (2005) 51–54.
- [12] J.R. Chien, G. Aletti, D.A. Bell, G.L. Keeney, V. Shridhar, L.C. Hartmann, Molecular pathogenesis and therapeutic targets in epithelial ovarian cancer, *J. Cell. Biochem.* 102 (2007) 1117–1129.
- [13] L. Dubeau, The cell of origin of ovarian epithelial tumours, *Lancet Oncol.* 9 (2008) 1191–1197.
- [14] K. Spiekermann, S. Biethahn, S. Wilde, W. Hiddemann, F. Alves, Constitutive activation of Stat transcription factors in acute myelogenous leukemia, *Eur. J. Haematol.* 67 (2001) 63–71.
- [15] N. Auersperg, M.M. Woo, C.B. Gilks, The origin of ovarian carcinomas: a developmental view, *Gynecol. Oncol.* 110 (2008) 452–454.
- [16] K. Shield, M.L. Ackland, N. Ahmed, G.E. Rice, Multicellular spheroids in ovarian cancer metastases: biology and pathology, *Gynecol. Oncol.* 113 (2009) 143–148.
- [17] K. Shield, C. Riley, M.A. Quinn, G.E. Rice, M.L. Ackland, N. Ahmed, Alpha2beta1 integrin affects metastatic potential of ovarian carcinoma spheroids by supporting disaggregation and proteolysis, *J. Carcinog.* 6 (2007) 11.
- [18] M.P. Ponnusamy, S.K. Batra, Ovarian cancer: emerging concept on cancer stem cells, *J. Ovarian Res.* 1 (2008) 4.
- [19] M.M. Gottesman, Mechanisms of cancer drug resistance, *Annu. Rev. Med.* 53 (2002) 615–627.

- [20] L. Zhang, J.R. Conejo-Garcia, D. Katsaros, P.A. Gimotty, M. Massobrio, G. Regnani, A. Makrigiannakis, H. Gray, K. Schlienger, M.N. Liebman, S.C. Rubin, G. Coukos, Intratumoral T cells, recurrence, and survival in epithelial ovarian cancer, *N. Engl. J. Med.* 348 (2003) 203–213.
- [21] A. Sica, T. Schioppa, A. Mantovani, P. Allavena, Tumour-associated macrophages are a distinct M2 polarised population promoting tumour progression: potential targets of anti-cancer therapy, *Eur. J. Cancer* 42 (2006) 717–727.
- [22] C. Lan, X. Huang, S. Lin, H. Huang, Q. Cai, T. Wan, J. Lu, J. Liu, Expression of M2-polarized macrophages is associated with poor prognosis for advanced epithelial ovarian cancer, *Technol. Cancer Res. Treat.* 12 (2013) 259–267.
- [23] I. Kryczek, M. Banerjee, P. Cheng, L. Vatan, W. Szeliga, S. Wei, E. Huang, E. Finlayson, D. Simeone, T.H. Welling, A. Chang, G. Coukos, R. Liu, W. Zou, Phenotype, distribution, generation, and functional and clinical relevance of Th17 cells in the human tumor environments, *Blood* 114 (2009) 1141–1149.
- [24] T. Kusuda, K. Shigemasa, K. Arihiro, T. Fujii, N. Nagai, K. Ohama, Relative expression levels of Th1 and Th2 cytokine mRNA are independent prognostic factors in patients with ovarian cancer, *Oncol. Rep.* 13 (2005) 1153–1158.
- [25] Y. Miyahara, K. Odunsi, W. Chen, G. Peng, J. Matsuzaki, R.F. Wang, Generation and regulation of human CD4⁺ IL-17-producing T cells in ovarian cancer, *Proc. Natl. Acad. Sci. U. S. A.* 105 (2008) 15505–15510.
- [26] T. Korn, E. Bettelli, M. Oukka, V.K. Kuchroo, IL-17 and Th17 cells, *Annu. Rev. Immunol.* 27 (2009) 485–517.
- [27] L.L. Chen, F. Ye, W.G. Lu, Y. Yu, H.Z. Chen, X. Xie, Evaluation of immune inhibitory cytokine profiles in epithelial ovarian carcinoma, *J. Obstet. Gynaecol. Res.* 35 (2009) 212–218.
- [28] T.J. Curiel, S. Wei, H. Dong, X. Alvarez, P. Cheng, P. Mottram, R. Krzysiek, K.L. Knutson, B. Daniel, M.C. Zimmermann, O. David, M. Burow, A. Gordon, N. Dhurandhar, L. Myers, R. Berggren, A. Hemminki, R.D. Alvarez, D. Emilie, D.T. Curiel, L. Chen, W. Zou, Blockade of B7-H1 improves myeloid dendritic cell-mediated antitumor immunity, *Nat. Med.* 9 (2003) 562–567.
- [29] W.J. Leonard, J.-X. Lin, Cytokine receptor signaling pathways, *J. Allergy Clin. Immunol.* 105 (2000) 877–888.
- [30] F. Burke, M. Relf, R. Negus, F. Balkwill, A cytokine profile of normal and malignant ovary, *Cytokine* 8 (1996) 578–585.
- [31] M. Nash, G. Ferrandina, M. Gordinier, A. Loercher, R. Freedman, The role of cytokines in both the normal and malignant ovary, *Endocr. Relat. Cancer* 6 (1999) 93–107.
- [32] L.A. O'Sullivan, C. Liongue, R.S. Lewis, S.E.M. Stephenson, A.C. Ward, Cytokine receptor signaling through the Jak/Stat/Socs pathway in disease, *Mol. Immunol.* 44 (2007) 2497–2506.
- [33] P.C. Heinrich, I. Behrmann, S. Haan, H.M. Hermanns, G. Muller-Newen, F. Schaper, Principles of interleukin (IL)-6-type cytokine signalling and its regulation, *Biochem. J.* 374 (2003) 1–20.
- [34] T. Kishimoto, Interleukin-6: from basic science to medicine – 40 years in immunology, *Annu. Rev. Immunol.* 23 (2005) 1–21.
- [35] V. Machelon, D. Emilie, A. Lefevre, F. Nome, I. Durand-Gasselien, J. Testart, Interleukin-6 biosynthesis in human preovulatory follicles: some of its potential roles at ovulation, *J. Clin. Endocrinol. Metab.* 79 (1994) 633–642.
- [36] M. Trouillas, C. Saucourt, B. Guillotin, X. Gauthereau, J.L. Taupin, J.F. Moreau, H. Boeuf, The IL-6 cytokine: towards adulthood, *Eur. Cytokine Netw.* 20 (2009) 51–62.
- [37] Y. Morikawa, Oncostatin M in the development of the nervous system, *Anat. Sci. Int.* 80 (2005) 53–59.
- [38] C. Liongue, C. Wright, A.P. Russell, A.C. Ward, Granulocyte colony-stimulating factor receptor: stimulating granulopoiesis and much more, *Int. J. Biochem. Cell Biol.* 41 (2009) 2372–2375.
- [39] L. Huang, C. Li, Leptin: a multifunctional hormone, *Cell Res.* 10 (2000) 81–92.
- [40] D. Kamimura, K. Ishihara, T. Hirano, IL-6 signal transduction and its physiological roles: the signal orchestration model, *Rev. Physiol. Biochem. Pharmacol.* 149 (2003) 1–38.
- [41] K. Imada, W.J. Leonard, The JAK-STAT pathway, *Mol. Immunol.* 37 (2000) 1–11.
- [42] J.S. Rawlings, K.M. Rosler, D.A. Harrison, The JAK/STAT signaling pathway, *J. Cell Sci.* 117 (2004) 1281–1283.
- [43] H. Yu, R. Jove, The Stats of cancer – new molecular targets come of age, *Nat. Rev. Cancer* 4 (2004) 97–105.
- [44] D.E. Levy, C. Lee, What does Stat3 do? *J. Clin. Invest.* 109 (2002) 1143–1148.
- [45] E.M. Dijkgraaf, M.J. Welters, J.W. Nortier, S.H. van der Burg, J.R. Kroep, Interleukin-6/interleukin-6 receptor pathway as a new therapy target in epithelial ovarian cancer, *Curr. Pharm. Des.* 18 (2012) 3816–3827.
- [46] D.R. Hodge, E.M. Hurt, W.L. Farrar, The role of IL-6 and STAT3 in inflammation and cancer, *Eur. J. Cancer* 41 (2005) 2502–2512.
- [47] N. Ahmed, F. Pansino, R. Clyde, P. Murthi, M.A. Quinn, G.E. Rice, M.V. Agrez, S. Mok, M.S. Baker, Overexpression of alpha(v)beta6 integrin in serous epithelial ovarian cancer regulates extracellular matrix degradation via the plasminogen activation cascade, *Carcinogenesis* 23 (2002) 237–244.
- [48] Y. Wang, L. Li, X. Guo, X. Jin, W. Sun, X. Zhang, R.C. Xu, Interleukin-6 signaling regulates anchorage-independent growth, proliferation, adhesion and invasion in human ovarian cancer cells, *Cytokine* 59 (2012) 228–236.
- [49] Y. Wang, X.L. Niu, Y. Qu, J. Wu, Y.Q. Zhu, W.J. Sun, L.Z. Li, Autocrine production of interleukin-6 confers cisplatin and paclitaxel resistance in ovarian cancer cells, *Cancer Lett.* 295 (2010) 110–123.
- [50] D.J. Kim, K.S. Chan, S. Sano, J. Digiovanni, Signal transducer and activator of transcription 3 (Stat3) in epithelial carcinogenesis, *Mol. Carcinog.* 46 (2007) 725–731.
- [51] S. Wormald, D.J. Hilton, Inhibitors of cytokine signal transduction, *J. Biol. Chem.* 279 (2004) 821–824.
- [52] V.M. Lauta, Interleukin-6 and the network of several cytokines in multiple myeloma: an overview of clinical and experimental data, *Cytokine* 16 (2001) 79–86.
- [53] S. Bellone, K. Watts, S. Cane, M. Palmieri, M.J. Cannon, A. Burnett, J.J. Roman, S. Pecorelli, A.D. Santin, High serum levels of interleukin-6 in endometrial carcinoma are associated with uterine serous papillary histology, a highly aggressive and chemotherapy-resistant variant of endometrial cancer, *Gynecol. Oncol.* 98 (2005) 92–98.
- [54] N. Songur, B. Kuru, F. Kalkan, C. Ozdilekan, H. Cakmak, N. Hize, Serum interleukin-6 levels correlate with malnutrition and survival in patients with advanced non-small cell lung cancer, *Tumori* 90 (2004) 196–200.
- [55] C. Belluco, D. Nitti, M. Frantz, P. Toppan, D. Basso, M. Plebani, M. Lise, J.M. Jessup, Interleukin-6 blood level is associated with circulating carcinoembryonic antigen and prognosis in patients with colorectal cancer, *Ann. Surg. Oncol.* 7 (2000) 133–138.
- [56] O. Altundag, K. Altundag, E. Gunduz, Interleukin-6 and C-reactive protein in metastatic renal cell carcinoma, *J. Clin. Oncol.* 23 (2005) 1044–1045.
- [57] L.H. Wei, M.L. Kuo, C.A. Chen, C.H. Chou, K.B. Lai, C.N. Lee, C.Y. Hsieh, Interleukin-6 promotes cervical tumor growth by VEGF-dependent angiogenesis via a STAT3 pathway, *Oncogene* 22 (2003) 1517–1527.
- [58] I. Garcia-Tunon, M. Ricote, A. Ruiz, B. Fraile, R. Paniagua, M. Royuela, IL-6, its receptors and its relationship with BCL-2 and BAX proteins in infiltrating and in situ human breast carcinoma, *Histopathology* 47 (2005) 82–89.
- [59] M.B. Nilsson, R.R. Langley, I.J. Fidler, Interleukin-6, secreted by human ovarian carcinoma cells, is a potent proangiogenic cytokine, *Cancer Res.* 65 (2005) 10794–10800.
- [60] P.M. Schwartzburd, Chronic inflammation as inductor of pro-cancer microenvironment: pathogenesis of dysregulated feedback control, *Cancer Metastasis Rev.* 22 (2003) 95–102.
- [61] J.M. Watson, J.L. Sensitaffar, J.S. Berek, O. Martinez-Maza, Constitutive production of interleukin 6 by ovarian cancer cell lines and by primary ovarian tumor cultures, *Cancer Res.* 50 (1990) 6959–6965.
- [62] S. Wu, K. Rodabaugh, O. Martinez-Maza, J. Watson, D. Silberstein, C. Boyer, W. Peters, J. Weinberg, J. Berek, R. Bast Jr., Stimulation of ovarian tumor cell proliferation with monocyte products including interleukin-1, interleukin-6, and tumor necrosis factor- α , *Am. J. Obstet. Gynecol.* 166 (1992) 997–1007.
- [63] C. Alberti, P. Pinciroli, B. Valeri, R. Ferri, A. Ditto, K. Umezawa, M. Sensi, S. Canevari, A. Tomasetti, Ligand-dependent EGFR activation induces the co-expression of IL-6 and PAI-1 via the NFkB pathway in advanced-stage epithelial ovarian cancer, *Oncogene* 31 (2012) 4139–4149.
- [64] M. Colomiere, A.C. Ward, C. Riley, M.K. Trenerry, D. Cameron-Smith, J. Findlay, L. Ackland, N. Ahmed, Cross talk of signals between EGFR and IL-6R through JAK2/STAT3 mediate epithelial-mesenchymal transition in ovarian carcinomas, *Br. J. Cancer* 100 (2009) 134–144.
- [65] S.K. Lutgendorf, A.Z. Weinrib, F. Penedo, D. Russell, K. DeGeest, E.S. Costanzo, P.J. Henderson, S.E. Sephton, N. Rohleder, J.A. Lucci III, S. Cole, A.K. Sood, D.M. Lubaroff, Interleukin-6, cortisol, and depressive symptoms in ovarian cancer patients, *J. Clin. Oncol.* 26 (2008) 4820–4827.
- [66] G. Scambia, U. Testa, P. Benedetti Panici, E. Foti, R. Martucci, A. Gadducci, A. Perillo, V. Facchini, C. Peschle, S. Mancuso, Prognostic significance of interleukin 6 serum levels in patients with ovarian cancer, *Br. J. Cancer* 71 (1995) 354–356.
- [67] J.S. Berek, C. Chung, K. Kaldi, J.M. Watson, R.M. Knox, O. Martinez-Maza, Serum interleukin-6 levels correlate with disease status in patients with epithelial ovarian cancer, *Am. J. Obstet. Gynecol.* 164 (1991) 1038–1042 (discussion 1042–1033).
- [68] R.L. Stone, A.M. Nick, I.A. McNeish, F. Balkwill, H.D. Han, J. Bottsford-Miller, R. Rupaimele, G.N. Armaiz-Pena, C.V. Pecot, J. Coward, M.T. Deavers, H.G. Vasquez, D. Urbauer, C.N. Landen, W. Hu, H. Gershenson, K. Matsuo, M.M. Shahzad, E.R. King, I. Tekedereli, B. Ozpolat, E.H. Ahn, V.K. Bond, R. Wang, A.F. Drew, F. Gushiken, D. Lamkin, K. Collins, K. DeGeest, S.K. Lutgendorf, W. Chiu, G. Lopez-Berestein, V. Afshar-Kharghan, A.K. Sood, Paraneoplastic thrombocytosis in ovarian cancer, *N. Engl. J. Med.* 366 (2012) 610–618.
- [69] M.S. Anglesio, J. George, H. Kulbe, M. Friedlander, D. Rischin, C. Lemeche, J. Power, J. Coward, P.A. Cowin, C.M. House, P. Chakravarty, K.L. Goringe, I.G. Campbell, Australian Ovarian Cancer Study, A. Okamoto, M.J. Birrer, D.G. Huntsman, A. de Fazio, S.E. Kalloger, F. Balkwill, C.B. Gilks, D.D. Bowtell, IL6-STAT3-HIF signaling and therapeutic response to the angiogenesis inhibitor sunitinib in ovarian clear cell cancer, *Clin. Cancer Res.* 17 (2011) 2538–2548.
- [70] K.S. Rath, H.M. Funk, M.C. Bowling, W.E. Richards, A.F. Drew, Expression of soluble interleukin-6 receptor in malignant ovarian tissue, *Am. J. Obstet. Gynecol.* 203 (2010) 230.e1–8.
- [71] N.H. Obata, K. Tamakoshi, K. Shibata, F. Kikkawa, Y. Tomoda, Effects of interleukin-6 on in vitro cell attachment, migration and invasion of human ovarian carcinoma, *Anticancer Res.* 17 (1997) 337–342.
- [72] Z. Duan, R. Foster, D.A. Bell, J. Mahoney, K. Wolak, A. Vaidya, C. Hampel, H. Lee, M.V. Seiden, Signal transducers and activators of transcription 3 pathway activation in drug-resistant ovarian cancers, *Clin. Cancer Res.* 12 (2006) 5055–5063.
- [73] R.T. Penson, K. Kronish, Z. Duan, A.J. Feller, P. Stark, S.E. Cook, L.R. Duska, A.F. Fuller, A.K. Goodman, N. Nikrui, K.M. MacNeill, U.A. Matulonis, F.I. Preffer, M.V. Seiden, Cytokines IL-1 β , IL-2, IL-6, IL-8, MCP-1, GM-CSF and TNF α in patients with epithelial ovarian cancer and their relationship to treatment with paclitaxel, *Int. J. Gynecol. Cancer* 10 (2000) 33–41.
- [74] Z. Duan, A.J. Feller, R.T. Penson, B.A. Chabner, M.V. Seiden, Discovery of differentially expressed genes associated with paclitaxel resistance using cDNA array technology: analysis of interleukin (IL) 6, IL-8, and monocyte chemotactic protein 1 in the paclitaxel-resistant phenotype, *Clin. Cancer Res.* 5 (1999) 3445–3453.
- [75] J. Coward, H. Kulbe, P. Chakravarty, D. Leader, V. Vassileva, D.A. Leinster, R. Thompson, T. Schioppa, J. Nemeth, J. Vermeulen, N. Singh, N. Avril, J. Cummings, E. Rexhepaj, K. Jirstrom, W.M. Gallagher, D.J. Brennan, I.A. McNeish, F.R. Balkwill, Interleukin-6 as a therapeutic target in human ovarian cancer, *Clin. Cancer Res.* 17 (2011) 6083–6096.

- [76] D. Duluc, Y. Delneste, F. Tan, M.P. Moles, L. Grimaud, J. Lenoir, L. Preisser, I. Anegon, L. Catala, N. Ifrah, P. Descamps, E. Gamelin, H. Gascan, M. Hebbat, P. Jeannin, Tumor-associated leukemia inhibitory factor and IL-6 skew monocyte differentiation into tumor-associated macrophage-like cells, *Blood* 110 (2007) 4319–4330.
- [77] H. Roca, Z.S. Varsos, S. Sud, M.J. Craig, C. Ying, K.J. Pienta, CCL2 and interleukin-6 promote survival of human CD11b⁺ peripheral blood mononuclear cells and induce M2-type macrophage polarization, *J. Biol. Chem.* 284 (2009) 34342–34354.
- [78] K. Takaishi, Y. Komohara, H. Tashiro, H. Ohtake, T. Nakagawa, H. Katabuchi, M. Takeya, Involvement of M2-polarized macrophages in the ascites from advanced epithelial ovarian carcinoma in tumor progression via Stat3 activation, *Cancer Sci.* 101 (2010) 2128–2136.
- [79] P. Allavena, A. Sica, G. Solinas, C. Porta, A. Mantovani, The inflammatory micro-environment in tumor progression: the role of tumor-associated macrophages, *Crit. Rev. Oncol. Hematol.* 66 (2008) 1–9.
- [80] T. Korn, M. Mitsdoerffer, A.L. Croxford, A. Awasthi, V.A. Dardalhon, G. Galileos, P. Vollmar, G.L. Stritesky, M.H. Kaplan, A. Waisman, V.K. Kuchroo, M. Oukka, IL-6 controls Th17 immunity in vivo by inhibiting the conversion of conventional T cells into Foxp3⁺ regulatory T cells, *Proc. Natl. Acad. Sci. U. S. A.* 105 (2008) 18460–18465.
- [81] T. Kato, H. Furumoto, T. Ogura, Y. Onishi, M. Irahara, S. Yamano, M. Kamada, T. Aono, Expression of IL-17 mRNA in ovarian cancer, *Biochem. Biophys. Res. Commun.* 282 (2001) 735–738.
- [82] L. Wang, T. Yi, M. Kortylewski, D.M. Pardoll, D. Zeng, H. Yu, IL-17 can promote tumor growth through an IL-6–Stat3 signaling pathway, *J. Exp. Med.* 206 (2009) 1457–1464.
- [83] A. Alshamsan, Induction of tolerogenic dendritic cells by IL-6-secreting CT26 colon carcinoma, *Immunopharmacol. Immunotoxicol.* 34 (2012) 465–469.
- [84] I. Kryczek, S. Wei, G. Zhu, L. Myers, P. Mottram, P. Cheng, L. Chen, G. Coukos, W. Zou, Relationship between B7-H4, regulatory T cells, and patient outcome in human ovarian carcinoma, *Cancer Res.* 67 (2007) 8900–8905.
- [85] C.W. Lo, M.W. Chen, M. Hsiao, S. Wang, C.A. Chen, S.M. Hsiao, J.S. Chang, T.C. Lai, S. Rose-John, M.L. Kuo, L.H. Wei, IL-6 trans-signaling in formation and progression of malignant ascites in ovarian cancer, *Cancer Res.* 71 (2011) 424–434.
- [86] E. Dimitriadis, C. Stoikos, Y.L. Tan, L.A. Salamonsen, Interleukin 11 signaling components signal transducer and activator of transcription 3 (STAT3) and suppressor of cytokine signaling 3 (SOCS3) regulate human endometrial stromal cell differentiation, *Endocrinology* 147 (2006) 3809–3817.
- [87] H.H. Nandurkar, L. Robb, C.G. Begley, The role of IL-11 in hematopoiesis as revealed by a targeted mutation of its receptor, *Stem Cells* 16 (Suppl. 2) (1998) 53–65.
- [88] T.M. Li, C.M. Wu, H.C. Huang, P.C. Chou, Y.C. Fong, C.H. Tang, Interleukin-11 increases cell motility and up-regulates intercellular adhesion molecule-1 expression in human chondrosarcoma cells, *J. Cell. Biochem.* 113 (2012) 3353–3362.
- [89] V. Lay, J. Yap, S. Sonderegger, E. Dimitriadis, Interleukin 11 regulates endometrial cancer cell adhesion and migration via STAT3, *Int. J. Oncol.* 41 (2012) 759–764.
- [90] T.L. Putoczki, S. Thiem, A. Loving, R.A. Busuttill, N.J. Wilson, P.K. Ziegler, P.M. Nguyen, A. Preaudet, R. Farid, K.M. Edwards, Y. Boglev, R.B. Luwor, A. Jamicki, D. Horst, A. Boussioutas, J.K. Heath, O.M. Sieber, I. Pleines, B.T. Kile, A. Nash, F.R. Greten, B.S. McKenzie, M. Ernst, Interleukin-11 is the dominant IL-6 family cytokine during gastrointestinal tumorigenesis and can be targeted therapeutically, *Cancer Cell* 24 (2013) 257–271.
- [91] C.L. Campbell, R. Guardiani, C. Ollari, B.E. Nelson, P.J. Quesenberry, T.M. Savarese, Interleukin-11 receptor expression in primary ovarian carcinomas, *Gynecol. Oncol.* 80 (2001) 121–127.
- [92] S. Wu, Y. Zhang, L. Xu, Y. Dai, Y. Teng, S. Ma, S.H. Ho, J.M. Kim, S.S. Yu, S. Kim, S. Song, Multicenter, randomized study of genetically modified recombinant human interleukin-11 to prevent chemotherapy-induced thrombocytopenia in cancer patients receiving chemotherapy, *Support. Care Cancer* 20 (2012) 1875–1884.
- [93] D.J. Hilton, N.A. Nicola, D. Metcalf, Specific binding of murine leukemia inhibitory factor to normal and leukemic monocytic cells, *Proc. Natl. Acad. Sci. U. S. A.* 85 (1988) 5971–5975.
- [94] D. Stejskal, V. Ruzicka, Cardiotrophin-1. Review, *Biomed. Pap. Med. Fac. Univ. Palacky Olomouc Czech Repub.* 152 (2008) 9–19.
- [95] K. Dhingra, A. Sahin, K. Emami, G.N. Hortobagyi, Z. Estrov, Expression of leukemia inhibitory factor and its receptor in breast cancer: a potential autocrine and paracrine growth regulatory mechanism, *Breast Cancer Res. Treat.* 48 (1998) 165–174.
- [96] P. Kellokumpu-Lehtinen, M. Talpaz, D. Harris, Q. Van, R. Kurzrock, Z. Estrov, Leukemia-inhibitory factor stimulates breast, kidney and prostate cancer cell proliferation by paracrine and autocrine pathways, *Int. J. Cancer* 66 (1996) 515–519.
- [97] T.M. Savarese, C.L. Campbell, C. McQuain, K. Mitchell, R. Guardiani, P.J. Quesenberry, B.E. Nelson, Coexpression of oncostatin M and its receptors and evidence for STAT3 activation in human ovarian carcinomas, *Cytokine* 17 (2002) 324–334.
- [98] C.H. Choi, J.J. Choi, Y.A. Park, Y.Y. Lee, S.Y. Song, C.O. Sung, T. Song, M.K. Kim, T.J. Kim, J.W. Lee, H.J. Kim, D.S. Bae, B.G. Kim, Identification of differentially expressed genes according to chemosensitivity in advanced ovarian serous adenocarcinomas: expression of GRIA2 predicts better survival, *Br. J. Cancer* 107 (2012) 91–99.
- [99] F. Ye, Y. Hu, W. Lu, C. Zhou, X. Xie, Expression of leukaemia inhibitory factor in epithelial ovarian carcinoma: correlation with clinical characteristics, *Histopathology* 53 (2008) 224–228.
- [100] P. Jeannin, D. Duluc, Y. Delneste, IL-6 and leukemia-inhibitory factor are involved in the generation of tumor-associated macrophage: regulation by IFN- γ , *Immunotherapy* 3 (2011) 23–26.
- [101] G. Dey, A. Radhakrishnan, N. Syed, J.K. Thomas, A. Nadig, K. Srikanth, P.P. Mathur, A. Pandey, S.K. Lin, R. Raju, T.S. Prasad, Signaling network of oncostatin M pathway, *J. Cell Commun. Signal.* 7 (2013) 103–108.
- [102] M.J. Gomez-Lechon, Oncostatin M: signal transduction and biological activity, *Life Sci.* 65 (1999) 2019–2030.
- [103] A.M. Douglas, G.A. Goss, R.L. Sutherland, D.J. Hilton, M.C. Berndt, N.A. Nicola, C.G. Begley, Expression and function of members of the cytokine receptor superfamily on breast cancer cells, *Oncogene* 14 (1997) 661–669.
- [104] M.K. Ganapathi, A.K. Weizer, S. Borsellino, R.M. Bukowski, R. Ganapathi, T. Rice, G. Casey, K. Kawamura, Resistance to interleukin 6 in human non-small cell lung carcinoma cell lines: role of receptor components, *Cell Growth Differ.* 7 (1996) 923–929.
- [105] S. Mori, K. Murakami-Mori, B. Bonavida, Oncostatin M (OM) promotes the growth of DU 145 human prostate cancer cells, but not PC-3 or LNCaP, through the signaling of the OM specific receptor, *Anticancer Res.* 19 (1999) 1011–1015.
- [106] Q. Li, J. Zhu, F. Sun, L. Liu, X. Liu, Y. Yue, Oncostatin M promotes proliferation of ovarian cancer cells through signal transducer and activator of transcription 3, *Int. J. Mol. Med.* 28 (2011) 101–108.
- [107] W.T. Watford, B.D. Hissong, J.H. Bream, Y. Kanno, L. Muul, J.J. O'Shea, Signaling by IL-12 and IL-23 and the immunoregulatory roles of STAT4, *Immunol. Rev.* 202 (2004) 139–156.
- [108] M. Del Vacchio, E. Bajetta, S. Canova, M.T. Lotze, A. Wesa, G. Parmiani, A. Anichini, Interleukin 12: biological properties and clinical applications, *Clin. Cancer Res.* 13 (2007) 4677–4685.
- [109] A.M. Wolf, H. Rumpold, D. Reimer, C. Marth, A.G. Zeimet, D. Wolf, High IL-12 p35 and IL-23 p19 mRNA expression is associated with superior outcome in ovarian cancer, *Gynecol. Oncol.* 118 (2010) 244–250.
- [110] N. Bozkurt, K. Yuce, M. Basaran, S. Gariboglu, F. Kose, A. Ayhan, Correlation of serum and ascitic IL-12 levels with second-look laparotomy results and disease progression in advanced epithelial ovarian cancer patients, *Int. J. Gynecol. Cancer* 16 (2006) 83–86.
- [111] K. Anwer, F.J. Kelly, C. Chu, J.G. Fewell, D. Lewis, R.D. Alvarez, Phase I trial of a formulated IL-12 plasmid in combination with carboplatin and docetaxel chemotherapy in the treatment of platinum-sensitive recurrent ovarian cancer, *Gynecol. Oncol.* 131 (2013) 169–173.
- [112] Q. Zhao, C. Wang, J. Zhu, L. Wang, S. Dong, G. Zhang, J. Tian, RNAi-mediated knock-down of cyclooxygenase2 inhibits the growth, invasion and migration of SaOS2 human osteosarcoma cells: a case control study, *J. Exp. Clin. Cancer Res.* 30 (2011) 26.
- [113] R.A. Kastelein, C.A. Hunter, D.J. Cua, Discovery and biology of IL-23 and IL-27: related but functionally distinct regulators of inflammation, *Annu. Rev. Immunol.* 25 (2007) 221–242.
- [114] J.L. Langowski, X. Zhang, L. Wu, J.D. Mattson, T. Chen, K. Smith, B. Basham, T. McClanahan, R.A. Kastelein, M. Olt, IL-23 promotes tumour incidence and growth, *Nature* 442 (2006) 461–465.
- [115] F. Pan, H. Yu, E.V. Dang, J. Barbi, X. Pan, J.F. Grosso, D. Jinasena, S.M. Sharma, E.M. McCadden, D. Getnet, C.G. Drake, J.O. Liu, M.C. Ostrowski, D.M. Pardoll, Eos mediates Foxp3-dependent gene silencing in CD4⁺ regulatory T cells, *Science* 325 (2009) 1142–1146.
- [116] Z. Zhang, B. Zhou, J. Zhang, Y. Chen, T. Lai, L. Yan, A. Liang, Y. Li, Y. Wang, Y. Chen, L. Zhang, M.R. Xi, Association of interleukin-23 receptor gene polymorphisms with risk of ovarian cancer, *Cancer Genet. Cytogenet.* 196 (2010) 146–152.
- [117] I.P. Touw, G.J. van de Geijn, Granulocyte colony-stimulating factor and its receptor in normal myeloid cell development, leukemia and related blood cell disorders, *Front. Biosci.* 12 (2007) 800–815.
- [118] A.D. Panopoulos, S.S. Watowich, Granulocyte colony-stimulating factor: molecular mechanisms of action during steady state and 'emergency' hematopoiesis, *Cytokine* 42 (2008) 277–288.
- [119] A.W. Roberts, G-CSF: a key regulator of neutrophil production, but that's not all! *Growth Factors* 23 (2005) 33–41.
- [120] A. Chakraborty, S.M. White, S.P. Lerner, Granulocyte colony-stimulating factor receptor signals for β 1-integrin expression and adhesion in bladder cancer, *Urology* 63 (2004) 177–183.
- [121] C.M. Gutschalk, C.C. Herold-Mende, N.E. Fusenig, M.M. Mueller, Granulocyte colony-stimulating factor and granulocyte-macrophage colony-stimulating factor promote malignant growth of cells from head and neck squamous cell carcinomas in vivo, *Cancer Res.* 66 (2006) 8026–8036.
- [122] G. Westphal, E. Niederberger, C. Blum, Y. Wollman, T.A. Knoch, W. Rebel, J. Debus, E. Friedrich, Erythropoietin and G-CSF receptors in human tumor cells: expression and aspects regarding functionality, *Tumori* 88 (2002) 150–159.
- [123] T. Brandstetter, E. Ninci, U. Falken, E. Wagner, R. Hess, T. Bauknecht, rhG-CSF affects genes involved in mitogen signalling and early gene expression in the ovarian cancer cell line HEY, *Int. J. Cancer* 75 (1998) 847–854.
- [124] E.B. Ninci, T. Brandstetter, I. Meinhold-Heerlein, H. Bettendorf, D. Sellin, T. Bauknecht, G-CSF receptor expression in ovarian cancer, *Int. J. Gynecol. Cancer* 10 (2000) 19–26.
- [125] T.M. Savarese, K. Mitchell, C. McQuain, C.L. Campbell, R. Guardiani, J. Wu, C. Ollari, F. Reale, B.E. Nelson, A. Chen, P.J. Quesenberry, Coexpression of granulocyte colony stimulating factor and its receptor in primary ovarian carcinomas, *Cancer Lett.* 162 (2001) 105–115.
- [126] T. Brandstetter, E. Ninci, H. Bettendorf, G. Perewusnyk, J. Stolte, D. Herchenbach, D. Sellin, E. Wagner, O.R. Kochli, T. Bauknecht, Granulocyte colony-stimulating factor (G-CSF) receptor gene expression of ovarian carcinoma does not correlate with G-CSF caused cell proliferation, *Cancer* 91 (2001) 1372–1383.
- [127] K. Munstedt, A. Hackethal, K. Eskef, I. Hrgovic, F.E. Franke, Prognostic relevance of granulocyte colony-stimulating factor in ovarian carcinomas, *Arch. Gynecol. Obstet.* 282 (2010) 301–305.
- [128] D.M. Spinner, T. Brandstetter, M. Kiechle-Schwarz, A. Du Bois, P. Angel, T. Bauknecht, c-Jun expression and growth stimulation in human ovarian carcinoma cell lines following exposure to cytokines, *Int. J. Cancer* 63 (1995) 423–427.

- [129] J. Kumar, F.W. Fraser, C. Riley, N. Ahmed, D.R. McCulloch, A.C. Ward, Granulocyte colony-stimulating factor receptor signalling via Janus kinase 2/signal transducer and activator of transcription 3 in ovarian cancer, *Br. J. Cancer* (2014), <http://dx.doi.org/10.1038/bjc.2013.673> (in press).
- [130] L.A. Tartaglia, The leptin receptor, *J. Biol. Chem.* 272 (1997) 6093–6096.
- [131] K. Nakashima, M. Narazaki, T. Taga, Leptin receptor (OB-R) oligomerizes with itself but not with its closely related cytokine signal transducer gp130, *FEBS Lett.* 403 (1997) 79–82.
- [132] E.C. Villanueva, M.G. Myers Jr., Leptin receptor signaling and the regulation of mammalian physiology, *Int. J. Obes. (Lond.)* 32 (Suppl. 7) (2008) S8–S12.
- [133] S.C. Chua, I.K. Koutras, L. Han, S.M. Liu, J. Kay, S.J. Young, W.K. Chung, R.L. Leibel, Fine structure of the murine leptin receptor gene: splice site suppression is required to form two alternatively spliced transcripts, *Genomics* 15 (1997) 264–270.
- [134] P. Somasundar, K.A. Frankenberry, H. Skinner, G. Vedula, D.W. McFadden, D. Riggs, B. Jackson, R. Vangilder, S.M. Hileman, L.C. Vona-Davis, Prostate cancer cell proliferation is influenced by leptin, *J. Surg. Res.* 118 (2004) 71–82.
- [135] N.K. Saxena, D. Sharma, X. Ding, S. Lin, F. Marra, D. Merlin, F.A. Anania, Concomitant activation of the JAK/STAT, PI3K/AKT, and ERK signaling is involved in leptin-mediated promotion of invasion and migration of hepatocellular carcinoma cells, *Cancer Res.* 67 (2007) 2497–2507.
- [136] N.K. Saxena, P.M. Vertino, F.A. Anania, D. Sharma, Leptin-induced growth stimulation of breast cancer cells involves recruitment of histone acetyltransferases and mediator complex to Cyclin D1 promoter via activation of Stat3, *J. Biol. Chem.* 282 (2007) 13316–13325.
- [137] D. Sharma, N.K. Saxena, P.M. Vertino, F.A. Anania, Leptin promotes the proliferative response and invasiveness in human endometrial cancer cells by activating multiple signal-transduction pathways, *Endocr. Relat. Cancer* 13 (2006) 629–640.
- [138] N. Yin, D. Wang, H. Zhang, X. Yi, X. Sun, B. Shi, H. Wu, G. Wu, X. Wang, Y. Shang, Molecular mechanisms involved in the growth stimulation of breast cancer cells by leptin, *Cancer Res.* 64 (2004) 5870–5875.
- [139] X. Hu, S.C. Juneja, N.J. Mailhe, M.P. Cleary, Leptin—a growth factor in normal and malignant breast cells and for normal mammary gland development, *J. Natl. Cancer Inst.* 94 (2002) 1704–1711.
- [140] D. Yan, D. Avtanski, N.K. Saxena, D. Sharma, Leptin-induced epithelial–mesenchymal transition in breast cancer cells requires beta-catenin activation via Akt/GSK3- and MTA1/Wnt1 protein-dependent pathways, *J. Biol. Chem.* 287 (2012) 8598–8612.
- [141] U. Wazir, W. Al Sarakbi, W.G. Jiang, K. Mokbel, Evidence of an autocrine role for leptin and leptin receptor in human breast cancer, *Cancer Genomics Proteomics* 9 (2012) 383–387.
- [142] M.R. Hoda, G. Theil, N. Mohammed, K. Fischer, P. Fornara, The adipocyte-derived hormone leptin has proliferative actions on androgen-resistant prostate cancer cells linking obesity to advanced stages of prostate cancer, *J. Oncol.* 2012 (2012) 280–386.
- [143] S. Loffler, G. Aust, U. Kohler, K. Spanel-Borowski, Evidence of leptin expression in normal and polycystic human ovaries, *Mol. Hum. Reprod.* 7 (2001) 1143–1149.
- [144] S. Uddin, R. Bu, M. Ahmed, J. Abubaker, F. Al-Dayel, P. Bavi, K.S. Al-Kuraya, Overexpression of leptin receptor predicts an unfavorable outcome in Middle Eastern ovarian cancer, *Mol. Cancer* 8 (2009) 74.
- [145] J.H. Choi, S.H. Park, P.C. Leung, K.C. Choi, Expression of leptin receptors and potential effects of leptin on the cell growth and activation of mitogen-activated protein kinases in ovarian cancer cells, *J. Clin. Endocrinol. Metab.* 90 (2005) 207–210.
- [146] A. Ptak, E. Kolaczowska, E.L. Gregoraszczuk, Leptin stimulation of cell cycle and inhibition of apoptosis gene and protein expression in OVCAR-3 ovarian cancer cells, *Endocrine* 43 (2013) 394–403.
- [147] C. Chen, Y.C. Chang, M.S. Lan, M. Breslin, Leptin stimulates ovarian cancer cell growth and inhibits apoptosis by increasing Cyclin D1 and Mcl-1 expression via the activation of the MEK/ERK1/2 and PI3K/Akt signaling pathways, *Int. J. Oncol.* 42 (2013) 1113–1119.
- [148] J.H. Choi, K.T. Lee, P.C. Leung, Estrogen receptor alpha pathway is involved in leptin-induced ovarian cancer cell growth, *Carcinogenesis* 32 (2011) 589–596.
- [149] M. Trikha, R. Corringham, B. Klein, J.F. Rossi, Targeted anti-interleukin-6 monoclonal antibody therapy for cancer: a review of the rationale and clinical evidence, *Clin. Cancer Res.* 9 (2003) 4653–4665.
- [150] P.M. Voorhees, Q. Chen, D.J. Kuhn, G.W. Small, S.A. Hunsucker, J.S. Strader, R.E. Corringham, M.H. Zaki, J.A. Nemeth, R.Z. Orlowski, Inhibition of interleukin-6 signaling with CNTO 328 enhances the activity of bortezomib in preclinical models of multiple myeloma, *Clin. Cancer Res.* 13 (2007) 6469–6478.
- [151] L. Wallner, J. Dai, J. Escara-Wilke, J. Zhang, Z. Yao, Y. Lu, M. Trikha, J.A. Nemeth, M.H. Zaki, E.T. Keller, Inhibition of interleukin-6 with CNTO328, an anti-interleukin-6 monoclonal antibody, inhibits conversion of androgen-dependent prostate cancer to an androgen-independent phenotype in orchiectomized mice, *Cancer Res.* 66 (2006) 3087–3095.
- [152] T.B. Dorff, B. Goldman, J.K. Pinski, P.C. Mack, P.N. Lara Jr., P.J. Van Veldhuizen Jr., D.I. Quinn, N.J. Vogelzang, I.M. Thompson Jr., M.H. Hussain, Clinical and correlative results of SWOG S0354: a phase II trial of CNTO328 (siltuximab), a monoclonal antibody against interleukin-6, in chemotherapy-pretreated patients with castration-resistant prostate cancer, *Clin. Cancer Res.* 16 (2010) 3028–3034.
- [153] T. Puchalski, U. Prabhakar, Q. Jiao, B. Berns, H.M. Davis, Pharmacokinetic and pharmacodynamic modeling of an anti-interleukin-6 chimeric monoclonal antibody (siltuximab) in patients with metastatic renal cell carcinoma, *Clin. Cancer Res.* 16 (2010) 1652–1661.
- [154] Y. Guo, J. Nemeth, C. O'Brien, M. Susa, X. Liu, Z. Zhang, E. Choy, H. Mankin, F. Hornicek, Z. Duan, Effects of siltuximab on the IL-6-induced signaling pathway in ovarian cancer, *Clin. Cancer Res.* 16 (2010) 5759–5769.
- [155] P. Garnero, E. Thompson, T. Woodworth, J.S. Smolen, Rapid and sustained improvement in bone and cartilage turnover markers with the anti-interleukin-6 receptor inhibitor tocilizumab plus methotrexate in rheumatoid arthritis patients with an inadequate response to methotrexate: results from a substudy of the multicenter double-blind, placebo-controlled trial of tocilizumab in inadequate responders to methotrexate alone, *Arthritis Rheum.* 62 (2010) 33–43.
- [156] J.E. Minturn, A.E. Evans, J.G. Villablanca, G.A. Yanik, J.R. Park, S. Shusterman, S. Groshen, E.T. Hellriegel, D. Bensen-Kennedy, K.K. Matthay, G.M. Brodeur, J.M. Maris, Phase I trial of lestaurtinib for children with refractory neuroblastoma: a new approaches to neuroblastoma therapy consortium study, *Cancer Chemother. Pharmacol.* 68 (2011) 1057–1065.
- [157] E. Leah, Clinical trials: phase III trial results for tofacitinib bring new oral DMARD therapy a step closer for patients with rheumatoid arthritis, *Nat. Rev. Rheumatol.* 8 (2012) 561.
- [158] A. Younes, J. Romaguera, M. Fanale, P. McLaughlin, F. Hagemeister, A. Copeland, S. Neelapu, L. Kwak, J. Shah, S. de Castro Faria, S. Hart, J. Wood, R. Jayaraman, K. Ethirajulu, J. Zhu, Phase I study of a novel oral Janus kinase 2 inhibitor, SB1518, in patients with relapsed lymphoma: evidence of clinical and biologic activity in multiple lymphoma subtypes, *J. Clin. Oncol.* 30 (2012) 4161–4167.
- [159] J. Schust, B. Sperl, A. Hollis, T.U. Mayer, T. Berg, Stattic: a small-molecule inhibitor of STAT3 activation and dimerization, *Chem. Biol.* 13 (2006) 1235–1242.
- [160] H.K. Bid, D. Oswald, C. Li, C.A. London, J. Lin, P.J. Houghton, Anti-angiogenic activity of a small molecule STAT3 inhibitor LLL12, *PLoS One* 7 (2012) e35513.
- [161] P.K. Kandala, S.K. Srivastava, Regulation of Janus-activated kinase-2 (JAK2) by diindolylmethane in ovarian cancer cells in vitro and in vivo, *Drug Discov. Ther.* 6 (2012) 94–101.
- [162] M. Hedvat, D. Huszar, A. Herrmann, J.M. Gozgit, A. Schroeder, A. Sheehy, R. Buettner, D. Proia, C.M. Kowolik, H. Xin, B. Armstrong, G. Beberitz, S. Weng, L. Wang, M. Ye, K. McEachern, H. Chen, D. Morosini, K. Bell, M. Alimzhanov, S. Ioannidis, P. McCoon, Z.A. Cao, H. Yu, R. Jove, M. Zinda, The JAK2 inhibitor AZD1480 potentially blocks Stat3 signaling and oncogenesis in solid tumors, *Cancer Cell* 16 (2009) 487–497.
- [163] Z. Duan, R.Y. Ames, M. Ryan, F.J. Hornicek, H. Mankin, M.V. Seiden, CDDO-Me, a synthetic triterpenoid, inhibits expression of IL-6 and Stat3 phosphorylation in multi-drug resistant ovarian cancer cells, *Cancer Chemother. Pharmacol.* 63 (2009) 681–689.
- [164] M. Saydmohammed, D. Joseph, V. Syed, Curcumin suppresses constitutive activation of STAT-3 by up-regulating protein inhibitor of activated STAT-3 (PIAS-3) in ovarian and endometrial cancer cells, *J. Cell. Biochem.* 110 (2010) 447–456.
- [165] L. Cai, G. Zhang, X. Tong, Q. You, Y. An, Y. Wang, L. Guo, T. Wang, D. Zhu, J. Zheng, Growth inhibition of human ovarian cancer cells by blocking STAT3 activation with small interfering RNA, *Eur. J. Obstet. Gynecol. Reprod. Biol.* 148 (2010) 73–80.
- [166] E.S. Lee, K.K. Ko, Y.A. Joe, S.G. Kang, Y.K. Hong, Inhibition of STAT3 reverses drug resistance acquired in temozolomide-resistant human glioma cells, *Oncol. Lett.* 2 (2011) 115–121.
- [167] A.A. Kiger, D.L. Jones, C. Schulz, M.B. Rogers, M.T. Fuller, Stem cell self renewal specified by Jak-Stat activation in response to a support cell cue, *Science* 294 (2001) 2542–2545.
- [168] Y. Yahata, Y. Shirakata, S. Tokumaru, K. Yamasaki, K. Sayama, Y. Hanakawa, M. Detmar, K. Hashimoto, Nuclear translocation of phosphorylated STAT3 is essential for vascular endothelial growth factor-induced human dermal microvascular endothelial cell migration and tube formation, *J. Biol. Chem.* 278 (2003) 40026–40031.
- [169] T. Wang, G. Niu, M. Kortylewski, L. Burdelya, K. Shain, S. Zhang, R. Bhattacharya, D. Gabrilovich, R. Heller, D. Coppola, W. Dalton, R. Jove, D. Pardoll, H. Yu, Regulation of the innate and adaptive immune responses by Stat-3 signaling in tumor cells, *Nat. Med.* 10 (2004) 48–54.
- [170] N.B. Davis, C.W. Ryan, W.M. Stadler, N.J. Vogelzang, A phase II study of nilutamide in men with prostate cancer after the failure of flutamide or bicalutamide therapy, *BJU Int.* 96 (2005) 787–790.
- [171] L.H. Wei, H. Baumann, E. Tracy, Y. Wang, A. Hutson, S. Rose-John, B.W. Henderson, Interleukin-6 trans signaling enhances photodynamic therapy by modulating cell cycling, *Br. J. Cancer* 97 (2007) 1513–1522.
- [172] I. Ray-Coquard, D. Paradiso, J.P. Guastalla, B. Leduc, F. Guichard, C. Martin, L. Chauvenet, Z. Haddad-Guichard, D. Lepille, H. Orfeuvre, H. Gautier, D. Castera, E. Pujade-Lauraine, Intensified dose of cyclophosphamide with G-CSF support versus standard dose combined with platinum in first-line treatment of advanced ovarian cancer: a randomised study from the GINECO group, *Br. J. Cancer* 97 (2007) 1200–1205.
- [173] Y. Yamada, M. Tomonaga, H. Fukuda, S. Hanada, A. Utsunomiya, M. Tara, M. Sano, S. Ikeda, K. Takatsuki, M. Kozuru, K. Araki, F. Kawano, M. Niimi, K. Tobinai, T. Hotta, M. Shimoyama, A new G-CSF-supported combination chemotherapy, LSG15, for adult T-cell leukaemia-lymphoma: Japan Clinical Oncology Group Study 9303, *Br. J. Haematol.* 113 (2001) 375–382.
- [174] K.M. Schmeler, S. Vadhan-Raj, P.T. Ramirez, S.M. Apte, L. Cohen, R.L. Bassett, R.B. Iyer, J.K. Wolf, C.L. Levenback, D.M. Gershenson, R.S. Freedman, A phase II study of GM-CSF and rIFN-gamma1b plus carboplatin for the treatment of recurrent, platinum-sensitive ovarian, fallopian tube and primary peritoneal cancer, *Gynecol. Oncol.* 113 (2009) 210–215.